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Antibody Stripping Buffer (For mIHC Fluorescence Kit)

Catalog Number: SLAR0002

Size: 30 mL

Format: Liquid, ready to use after warming

Storage: Room temperature (RT); stable for 12 months

Application: antibody elution in TSA-based multiplex immunofluorescence

DESCRIPTION

Antibody Stripping Buffer is a ready-to-use (after warming) elution reagent designed for multiplex immunofluorescence (mIHC / TSA) workflows. After each round of TSA staining, the antibody deposited on the section must be removed before the next primary antibody is applied; this buffer strips the bound antibody under mild conditions while leaving the covalently deposited tyramide fluorophore in place, so that several targets can be labelled sequentially on the same slide. It is recommended for frozen sections, cell smears, cell climbing slices and fragile tissues such as decalcified bone; for paraffin sections, heat retrieval in EDTA or citrate buffer is the preferred stripping method.

SPECIFICATIONS

Catalog Number	SLAR0002
Product Name	Antibody Stripping Buffer (For mIHC Fluorescence Kit)
Format	Liquid (ready to use after warming to 37℃)
Size	30 mL
Storage	Room temperature (RT); stable for 12 months
Applications	TSA-based multiplex immunofluorescence (mIHC)
Sample type	Frozen sections, cell smears, cell climbing slices, fragile tissue; paraffin sections optional
Intended Use	For research use only

PROTOCOL

1. Warm the buffer

Warm the buffer to 37℃ until it is completely dissolved and free of precipitate before each use.

2. Apply to the section

After the TSA fluorescence staining step and PBS washes (3 × 5 min), apply an appropriate volume of pre-warmed buffer to cover the tissue.

3. Incubate

Incubate for 5-20 min at 37℃, then discard the buffer.

4. Repeat

Apply fresh buffer once more to cover the tissue and incubate for another 5-20 min at 37℃, then discard.

5. Wash

Wash the slide with PBS (pH 7.4) for 5 min, repeated 3 times, and continue with the next round of primary antibody incubation.

NOTES

1. Stripping efficiency depends on section thickness, stripping temperature and time, and the primary antibody used; adjust the incubation time accordingly.
2. If stripping is incomplete, lower the primary antibody concentration or move that antibody to the last round.
3. The buffer flows off the section easily; replenish it promptly if it runs off.
4. Make sure the reagent is completely dissolved before use; if precipitate is present, warm at 37℃ until clear.
5. Preferentially use this buffer for cell smears, cell climbing slices, frozen sections and fragile bone tissue; for paraffin sections, EDTA or citrate heat retrieval is preferred.
6. The buffer is slightly acidic; over-stripping (too long or too hot) can reduce antigen recognition and weaken DAPI nuclear staining.
7. Do not use this buffer on unfixed sections.

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.